



Collecting Samples

It is very important to get a representative sample .

The methods for doing this vary with the type of sample that is being collected. The general methods for collecting representative samples are described below. Try to follow these suggestions as closely as possible if you have any questions, please call (919)-688-4804 or contact us by email at info@dioxins.com.

In addition to following the instructions we suggest preparing a detailed account of how the samples were collected. This information may be useful in the future should there be any question about the accuracy and validity of the sampling methods.

General Information:

If at all possible, samples should be collected into clean glass containers and must be secured with tight lids. Samples may be collected by using the sample jars as scoops and then sealing the jars. Please do not scoop sample material into jars using the same scoop. Each sample collection procedure should use clean (and not previously used) collection instruments.

All food and tissue samples should be shipped refrigerated (in insulated containers with non-water freezer packs). Soil, feed, and other samples not likely to spoil do not require refrigerated shipping. A minimum of 10 g of sample material is required for analysis. Additional 10 g amounts of sample material provide immediate opportunities for retesting in the event of unforeseen circumstances.

Please use sturdy packaging when shipping samples. Samples should be wrapped or surrounded with sufficient protective materials (bubble wrap, peanuts, Styrofoam, etc.) to protect from breakage. If you need sample collection glassware, freezer packs or shipping materials, please contact us.

If you are using a shipping container from our office, please place any enclosed freezer packs in your freezer upon arrival. Keep the caps on the bottles until you are ready to add the samples. After each sample has been collected, replace the cap securely and apply a completed, signed seal of custody across the cap. Fill out a sample identification label as to date and time of collection, who collected the sample and an identification of the sample. For your records, note which sample is associated with which bar code number on the sample identification label. Blank adhesive mailing labels can be used for this.

Xenobiotic Detection Systems, Inc.

"Dioxin Bioassays"



1601 East Geer St., Suite S
Durham, NC 27704
Phone: (919) 688-4804
888-DIOXINS (toll free)
Fax: (919) 688-4404
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Email: info@dioxins.com

Water Samples:

If the water is being collected from the tap, turn on the cold water and let it run for at least a minute. After the water has been running for a minute, collect the sample making sure not to touch the lip of the bottle to anything while the cap is off. Fill the bottle two-thirds to three-fourths full.

Surface Water Samples:

Try to get a representative sample. Attempt to exclude particulate material unless they are considered to be a representative part of the sample. Particulate materials will cause heterogeneity of the sample and may cause difficulty in interpreting results. When feasible, samples collected from the center of the body of water are preferable. Fill the bottle two-thirds to three-fourths full.

Blood samples:

CALUX[®] analyses can be performed on serum blood or whole blood. Samples should be withdrawn into glass "vacutainers" or similar glass receptacles. Please do not use "vacutainers" with serum separator material. A minimum of 10 cc of sample is required. A sample of 20 cc is preferred. A sample of 20 cc can be collected in a single 20 cc "vacutainer" tube or two 10 cc tubes. No additional preparation is required.

All blood samples should be immediately refrigerated. Shipping should be via next day delivery. Pack samples in styrofoam or similar insulated container with protective packaging to prevent breakage. All blood samples should also be enclosed in an additional container to prevent leakage should the original vial break. Include sufficient non-water freezer packs to keep samples refrigerated for length of shipment. If samples are not sent immediately to XDS, they may be frozen and shipped to XDS later using dry ice.

Returning the Samples:

If you are using Xenobiotic sample shipping materials, repack the package as it was sent to you.

If the samples need refrigeration, place an ice pack on the bottom of the cooler. Then set the four bottles (if using XDS packaging) upright on top of the ice pack and place the second ice pack between the bottles. Place the plastic tube of water between the bottles and the side of the cooler. Replace the styrofoam packing material around the bottles to help to secure them. All refrigerated samples should be shipped for next day delivery.

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Seal the container and mail to:

Xenobiotic Detection Systems, Inc.
1601 E. Geer St., Suite S
Durham, NC 27704
USA

Phone (919)-688-4804

Notification:

We would appreciate receiving an email notice of your shipping samples to us, containing the number of samples and sample material, shipping company, expected arrival date at XDS and the tracking number. Please send this to info@dioxins.com.